First hydroxamic seconucleoside derivatives

Vrček, Valerije; Čaplar, Vesna; Uršić, Stanko

Source / Izvornik: Croatica Chemica Acta, 1998, 71, 119 - 124

Journal article, Published version Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

Permanent link / Trajna poveznica: https://urn.nsk.hr/urn:nbn:hr:163:496056

Rights / Prava: In copyright/Zaštićeno autorskim pravom.

Download date / Datum preuzimanja: 2024-05-05



Repository / Repozitorij:

Repository of Faculty of Pharmacy and Biochemistry University of Zagreb



ISSN 0011-1643 CCA-2482 Preliminary Communication

First Hydroxamic Seconucleoside Derivatives

Valerije Vrček,^a Vesna Čaplar,^b and Stanko Uršić^a

^aFaculty of Pharmacy and Biochemistry, University of Zagreb, A. Kovačića 1, 10001 Zagreb, Croatia (e-mail: stu@nana.pharma.hr)

^bLaboratory of Supramolecular and Nucleoside Chemistry, Rugjer Bošković Institute, P.O.B. 1016, 10001 Zagreb, Croatia

Received May 21, 1997; revised September 15, 1997; accepted September 19, 1997

Nucleoside analogues possessing hydroxamate moiety were prepared from secouridine derivatives oxidized at C(2')-position. 3',5'-O-Isopropylidene-2',3'-secouridine was oxidized with CrO_3 /py complex to the carboxylate level and with DMSO/DCC to the aldehyde level. Both compounds (precursors) gave C(2')-nucleoside hydroxamic acids in the reaction with hydroxylamine or nitrosobenzene in the presence of Fe^{3+} ions, respectively.

A number of hydroxamic acid analogues have been shown to inhibit DNA synthesis by inactivating the enzyme ribonucleotide reductase (RNR). 1-4 This metaloenzyme catalyzes the conversion of (ribo)nucleotides to deoxy(ribo)nucleotides and is therefore a potential target for the development of anticancer agents.⁵⁻⁸ Hydroxamic acid moiety, R-CONHOH, is found to be the essential pharmacophore in the hydroxyurea, a clinically useful inhibitor of ribonucleotide reductase.9 A variety of nucleoside analogues are also active as inhibitors of ribonucleotide reductase, 10-12 following the inhibition mechanism similar to that proposed for hydroxamates. 13-15 These results have initiated our interest in the design and synthesis of the nucleoside analogues incorporating hydroxamate moiety. A similiar concept was devised by Farr et al. 16 Their compounds inhibited RNR activity, but were 10-fold less potent than hydroxyurea. We, therefore, rationalize that hydroxamate-nucleoside analogues which more closely resemble the RNR substrates should be prepared. Moreover, recent reports^{17–19} that hydroxamate compounds increase the potency of nucleosides against HIV-1 give an additional importance to derivatives combining the structural fea120 v. vrček *et al.*

tures of the above compounds. To prepare hydroxamic-nucleoside derivatives, we have used secouridine-2',3'-dialdehyde as the starting material. We have already reported preliminary results on the reaction of nitrosobenzene with 2',3'-secouridine dialdehyde giving hydroxamic-nucleoside product.²⁰ It is shown that this reaction does not proceed in the absence of ferric ion under the conditions employed. In the presented work, we also prepared monoaldehyde form **3** of 2',3'-secouridine **1** (Scheme 1) in order to prevent the formation of cyclic hemiacetals of dialdehyde monohydrate.^{21,22}

Scheme 1. a) acetone, H⁺, RT, 2 h; b) DMSO/benzene/pyridine, TFA and DCC, RT, overnight; c) Fe³⁺, H⁺, PhNO in acetone, RT, 6 h.

3',5'-O-Isopropylidene-2',3'-seconucleoside **2** was obtained according to the procedure described by Jones $et\ al.^{23}$ The oxidation of **2** to the aldehyde level was performed by dimethylsulfoxide (DMSO) and dicyclohexylcarbodimide (DCC) in the presence of pyridinium trifluoroacetate (PTFA).²⁴ The aldehyde product **3** was obtained in 60% yield. In the NMR spectra of **3** the signals of aldehyde proton at 10.30 ppm and aldehyde carbon at 194.20 ppm were present, although the compound exists predominately as hydrate **3a**. The test with dinitrophenylhydrazine was positive.

In the reaction of aldehyde **3** with nitrosobenzene, we have observed in UV spectrum the appearance of the highly characteristic peak at 520 nm due to the formation of the iron(III)-mono-*N*-phenylhydroxamate complex **4**. The reaction was promoted both with proton and Fe³⁺ ions. Anyway, the presence of Fe³⁺ in the reaction mixture made it difficult to isolate a free hydroxamic-nucleoside conjugate from complex product **4**. This is due to the known property of hydroxamates to form a strong complex with Fe³⁺.^{25,26} To circumvent this difficulty, we have applied a different approach to the synthesis of hydroxamic-nucleoside derivatives (Scheme 2).

Scheme 2. a) CrO_3 /pyridine complex in CH_2Cl_2 -DMF, AcOAc, t-butanol, RT, 20 h; b) NH_2OH in MeOH, RT, 3 h; c) 40% AcOH, RT, 3 h.

One-step oxidation of primary alcohol **2** to the corresponding carboxylic tert-butyl ester $\mathbf{5}^{27}$ was performed using chromium trioxide-pyridine complex²⁸ in dichloromethane-dimethylformamide (4:1) in the presence of tert-butanol and acetic anhydride (60% yield). 10% of the starting compound and 15% of acetylated by-product, 2'-acetyl-3',5'-O-isopropylidene-2',3'-secouridine (**6**), were also isolated from the reaction mixture. Omission of dimethylformamide from the reaction mixture led to poorer solubility of the oxidizing complex and consequently to slower oxidation. Therefore, in dichloromethane, the yield of oxidized product **5** was diminished (26%) and the ratio of acetylated product **6** doubled.

5'-O-Trityl-2',3'-secouridine **7** (Scheme 3.) could not be oxidized to tert-butyl carboxylates at C(2') and/or C(3') position by this method, presumably because of the complex cyclic hemiacetal structures of intermediary dialdehydes. The only isolated products were the acetylated starting compound, 2',3'-di-O-acetyl-2',3'-secouridine (8), and 1-[2(S)-trityloxymethyl-3(R,S)-acetoxy-1,4-dioxan-6(R)-yl]uracil (9). 1,4-Dioxacyclohexane structure **9** is a diastereomeric mixture resulting from acetylation of the previously formed intramolecular aldehyde hemiacetal. Also, an attempt at oxidation of 5'-O-trityl-2',3'-secouridine dialdehyde **10** resulted only in a complex mixture of diastereoisomeric cyclic acetals.

The method used for the preparation of 2'-hydroxamate derivative 11 (Scheme 2.) is similar to that described for the synthesis of the first sugar-

122 v. vrček $\it et al.$

C(2') and/or C(3') tert - butyl carboxylates

Scheme 3. a) CrO₃/pyridine complex in CH₂Cl₂-DMF, AcOAc, t-butanol, RT, 20 h.

hydroxamic acid.²⁹ Solution of sodium methoxide was added to hydroxylamine hydrochloride to prepare a free hydroxylamine. After removing sodium chloride by filtration hydroxylamine was reacted with *tert*-butyl-3',5'-O-isopropylidene-2',3'-secouridine-2'-carboxylate **5** giving 3',5'-O-isopropylidene-2',3'-secouridine-2'-hydroxamic acid **11** (65% yield). Deprotection was achieved in 40% acetic acid for 3 hours at room temperature giving hydroxamic-nucleoside derivative **12**³⁰ (65% yield). As far as we know, it is the first seconucleoside analogue incorporating hydroxamic moiety. We are continuing investigations on hydroxamic-nucleoside analogues with other nucleobases. Metal coordination properties and biological testing of compounds described will be reported in due course.

REFERENCES AND NOTES

- J. C. Swarts, M. A. S Aquino, J. Y. Han, K. Y. Lam, and A. G. Sykes, Biochim. Biophys. Acta Protein Struct. Mol. Enzymol. 1247 (1995) 215–224.
- 2. I. K. Larsen, B.-M. Sjöberg, and L. Thelander, Eur. J. Biochem. 125 (1982) 75–81.
- 3. B. van't Riet, G. L. Wampler, and H. L. Elford, J. Med. Chem. 22 (1979) 589-591.
- 4. H. L. Elford, G. L. Wampler, and B. van't Riet, Cancer Res. 39 (1979) 844-851.
- 5. P. Reichard, Science 260 (1993) 1773-1777.
- 6. J. Stubbe, Adv. Enzymol. Relat. Areas Mol. Biol. 63 (1990) 349–374.
- 7. J. Stubbe and W. A. van der Donk, Chem. Biol. 2 (1995) 793–801.
- 8. M. N. Prichard and C. Shipman, Chemotherapy 41 (1995) 384–395.
- 9. S. P. Gupta, Chem. Rev. **94** (1994) 1507–1551.
- 10. K. C. Xie and W. Plunkett, Cancer Res. **56** (1996) 3030–3037.
- 11. T. E. Lehmann and A. Berkessel, J. Org. Chem. 62 (1997) 302–309.
- 12. S. Salowe, J. M. Bolliger, M. Ator, J. Stubbe, J. McCracken, J. Peisach, M. C. Samano, and M. J. Robins, *Biochemistry* **32** (1993) 12749–12760.
- 13. M. J. Robins, Z. Guo, and S. F. Wnuk, J. Am. Chem. Soc. 119 (1997) 3637–3638.
- R. Eliasson, E. Pontis, F. Eckstein, and P. Reichard, J. Biol. Chem. 265 (1994) 26116–26120.

- 15. R. Ingemarson and L. Thelander, Biochemistry 35 (1996) 8603–8609.
- R. A. Farr, P. Bey, P. S. Sunkara, and B. J. Lippert, J. Med. Chem. 32 (1989) 1879– 1885.
- S. D. Malley, J. M. Grange, F. Hamedi-Sangsari, and J. R. Vila, *Proc. Natl. Acad. Sci. USA* 91 (1994) 11017–11021.
- F. Lori, A. Malykh, A. Cara, D. Sun, J. N. Weinstein, J. Lisziewicz, and R. C. Gallo, Science 266 (1994) 801–805.
- W. -Y. Gao, H. Mitsuya, J. S. Driscoll, and D. G. Johns, *Biochem. Pharmacol.* 50 (1995) 274–276.
- 20. S. Uršić, B. Nigović, V. Vrček, and V. Pilepić, *Tetrahedron Lett.* **36** (1995) 9547–9550.
- O. Howarth, A. S. Jones, R. T. Walker, and P. G. Wyatt, J. Chem. Soc. Perkin Trans. II (1984) 261–265.
- J. P. Neenan, S. M. Opitz, C. L. Cooke, M. A. Ussery, T. C. Morrill, and L. M. Eckel, *Bioorg. Med. Chem. Lett.* 6 (1996) 1381–1386.
- 23. A. S. Jones, M. J. McClean, H. Tanaka, R. T. Walker, J. Balzarini, and E. de Clercq, *Tetrahedron* 41 (1985) 5965–5972.
- 24. K. E. Pfitzner and J. G. Moffatt, J. Am. Chem. Soc. 87 (1965) 5670-5678.
- B. F. Matzanke, G. Mueler-Matzanke, and K. N. Raymond, Siderophore Mediated Iron Transport; Chemistry, Biology and Physical Properties, in: T. M. Loehr, H. B. Gray, and A. B. Lever (Eds.), Physical Bioorganic Chemistry, VCH Publishers, New York, 1989.
- 26. M. J. Miller and F. Malouin, Acc. Chem. Res. 25 (1993) 241–252.
- 27. 71 H-NMR (CDCl₃) δ /ppm: 9.0 (bs, 1H, NH), 7.51 (d, 1H, J = 8.13 Hz, 6-H), 6.10 (s, 1H, 1'-H), 5.82 (d, 1H, J = 8.14 Hz, 5-H), 4.09–3.70 (m, 5H, 3'-H₂, 4'-H, 5'-H₂), 1.49 (s, 9H, t-Bu), 1.42 and 1.41 (2s, 2x3H, iPr). 13 C-NMR (CDCl₃) δ /ppm: 164.80 (C-2'), 163.42 (C-4), 150.66 (C-2), 140.52 (C-6), 102.82 (C-5), 98.31 (CMe₂), 84.24 (CMe₃), 79.94 (C-1'), 71.60 (C-4'), 61.76 and 61.70 (C-3' and C-5'), 27.51 (CMe₃), 24.16 and 22.20 (CMe₂). Anal. Calcd. for C₁₆H₂₄N₂O₇ (M_r = 356.38): C 53.93, H 6.79, N 7.86%; found: C 53.98, H 7.03, N 7.71%.
- 28. E. J. Corey and B. Samuelsson, J. Org. Chem. 49 (1984) 4735–4735.
- 29. S. Uršić, B. Zorc, V. Pilepić, and D. Vikić-Topić, Croat. Chem. Acta 65 (1992) 851–857.
- 30. $^{1}\text{H-NMR}$ (DMSO-d₆) δ/ppm : 7.52 (d, 1H, J = 8.0 Hz, 6-H), 6.04 (s, 1H, 1'-H), 5.61 (d, 1H, J = 8.0 Hz, 5-H), 4.0–4.5 (m, NHOH, 2x OH), 3.55–3.33 (m, 5H, 3'-H₂, 4'-H, 5'-H₂). $^{13}\text{C-NMR}$ (DMSO-d₆) δ/ppm : 163.73 (C-2'), 161.65 (C-4), 151.29 (C-2), 142.48 (C-6), 101.49 (C-5), 81.92 and 81.40 (C-4' and C-1'), 60.77 and 60.58 (C-3' and C-5'). Anal. Calcd. for C $_{9}\text{H}_{13}\text{N}_{3}\text{O}_{7}$ (M $_{r}$ = 275.22): C 39.28, H 4.76, N 15.27%; found: C 39.33, H 4.75, N 15.30%.

124 V. VRČEK *ET AL*.

SAŽETAK

Prvi derivati hidroksamskih sekonukleozida

Valerije Vrček, Vesna Čaplar i Stanko Uršić

Nukleozidni derivati s hidroksamskom skupinom pripravljeni su iz sekouridina oksidiranog na položaju C(2'). 3',5'-O-Izopropiliden-2',3'-sekouridin oksidiran je s pomoću kompleksa CrO₃/piridin do karboksilata, a pomoću DMSO/DCC do aldehida. Karboksilatni derivat u reakciji s hidroksilaminom, odnosno aldehidni derivat u reakciji s nitrozobenzenom u prisutnosti iona Fe³⁺ daje C(2')-hidroksamski nukleozid.